2),

5′-TAT GGATCC TCAGCTGCAAATGAGGG-3′(26 mer; SEQ ID NO: 3),

5'-GTGGAATTCATGAAGAAAGAGATGATCATG-3′(30 mer; SEQ ID NO: 4),

5′-TATGGTACCTCAGCCGTCCGTGCTGCTT-3' (28 mer; SEQ ID NO: 5),

5'-GCGAAGCTTTGGAGAGTGGCATGAAGAAA-3′ (29 mer; SEQ ID NO: 6),

5'-TATGGATCCAACCATTCAACATGGTGGAC-3'(29 mer; SEQ ID NO: 7).

Please replace paragraph 125 with the following substitute paragraph.

The crude 29 mer 5'-GCGAAGCTTTGGAGAGTGGCATGAAGAAA-3′ (SEQ ID NO: 6) above was assembled on an $\mathbf{A_f}$-support and cleaved with 2M ammonia in methanol (20°C, 1hour), and finally deblocked by adding an equal amount of 32% ammonium hydroxide and heating at 55°C for an additional 4 hours. The structures of the long oligomers were confirmed by electrospray ionization mass spectra after chromatographic purification.

Please replace paragraph 126 with the following replacement paragraph.

Also, oligonucleotide phosphorothioate 5'-TGGCGTCTTCCATTT-3' (SEQ ID NO: 1) was synthesized on a commercial **T**-bound support and an $\mathbf{A_f}$ support following recommended protocols. No differences in coupling efficiency (>98% as determined from trityl assay) were detected between support $\mathbf{A_f}$ and the **T**-support. The support bound oligonucleotide phosphorothioate was cleaved and deblocked as described above and then analyzed by ion exchange HPLC. Analysis of the crude oligonucleotide phosphorothioate prepared on the $\mathbf{A_f}$ support indicated that the oligomer prepared on the universal support according to the invention was identical to the phosphoprothioate prepared on a conventional **T**-bound solid phase.

Please enter the enclosed paper copy of the Sequence Listing after the claims.